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August 10, 2000

EP00/5150

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APPLICATION NUMBER: 09/326,501

FILING DATE: June 04, 1999

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PTO-SB-05 (8-95)
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NEW UTILITY PATENT APPLICATION TRANSMITTAL <small>(to be used for new applications only)</small>	Attorney Docket Number	1.1.114
	First Named Inventor	E. Martinez Force
	Total Pages in this Submission	18

APPLICATION ELEMENTS	ACCOMPANYING APPLICATION PARTS						
<p>Notice: Checklist items mentioned under Application Elements section construct a new utility patent application. Please refer to MPEP Sections 506, 601, (37CFR 1.77, 1.53, 35 USC 111, 112, 113) for detailed explanation regarding completeness of an original patent application.</p> <p>1. <input type="checkbox"/> Fee Transmittal Form (prescribed filing fee(s))</p> <p>2. Specification</p> <p><input checked="" type="checkbox"/> Title of the Invention</p> <p><input type="checkbox"/> Cross References to Related Applications (if applicable)</p> <p><input type="checkbox"/> Statement Regarding Federally-sponsored Research/Development (if applicable)</p> <p><input type="checkbox"/> Reference to Microfiche Appendix (if applicable)</p> <p><input checked="" type="checkbox"/> Background of the Invention</p> <p><input checked="" type="checkbox"/> Brief Summary of the Invention</p> <p><input checked="" type="checkbox"/> Brief Description of the Drawings (if drawings filed)</p> <p><input checked="" type="checkbox"/> Detailed Description</p> <p><input checked="" type="checkbox"/> Claim or Claims</p> <p><input type="checkbox"/> Abstract of the Disclosure</p> <p>3. <input checked="" type="checkbox"/> Drawing(s) (when necessary as prescribed by 35 USC 113)</p> <p>4. <input type="checkbox"/> Executed Declaration</p> <p>5. Genetic Sequence Submission (if applicable, all must be included)</p> <p><input type="checkbox"/> Paper Copy</p> <p><input type="checkbox"/> Computer Readable Copy</p> <p><input type="checkbox"/> Statement Verifying Identical Paper and Computer Readable Copy</p>	<p>6. <input type="checkbox"/> Assignment Papers</p> <p>7. <input type="checkbox"/> Certified Copy of Priority Document(s) (if foreign priority is claimed)</p> <p>8. <input type="checkbox"/> Computer Program in Microfiche</p> <p>9. <input type="checkbox"/> English Translation Document (if applicable)</p> <p>10. <input type="checkbox"/> Information Disclosure Statement (PTO-1449) <input type="checkbox"/> Copies of IDS Citations</p> <p>11. <input type="checkbox"/> Petition Checklist and Accompanying Petition</p> <p>12. <input type="checkbox"/> Preliminary Amendment</p> <p>13. <input type="checkbox"/> Proprietary Information</p> <p>14. <input checked="" type="checkbox"/> Return Receipt Postcard</p> <p>15. <input type="checkbox"/> Small Entity Statement</p> <p>16. <input type="checkbox"/> Additional Enclosures (please identify below):</p> <p>SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT</p> <table border="1"><tr><td>Firm or Individual name</td><td>Garst Seed Company 33,762 Dana S. Rewoldt</td></tr><tr><td>Signature</td><td><i>Dana Rewoldt</i></td></tr><tr><td>Date</td><td>June 4, 1999</td></tr></table>	Firm or Individual name	Garst Seed Company 33,762 Dana S. Rewoldt	Signature	<i>Dana Rewoldt</i>	Date	June 4, 1999
Firm or Individual name	Garst Seed Company 33,762 Dana S. Rewoldt						
Signature	<i>Dana Rewoldt</i>						
Date	June 4, 1999						

FOR OFFICIAL USE ONLY					
Application Number			Class	Independent Claims	
Date of Receipt	Application Type		GAU	Total Claims	
	Filing Date		Foreign Filing License?	Drawing Sheets	
	Small Entity		Foreign Address?	Special Handling?	

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09/326501

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PATENT

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

Transmitted herewith for filing is the patent application of
Inventor(s): Enrique Martinez Force, Spanish, Lamacque de Novoa, 9 3°A,
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de la Concepcion, Sevilla, Spain

CERTIFICATION UNDER 37 C.F.R. 1.10⁴
(Express Mail label number is mandatory.)
(Express Mail certification is optional.)

I hereby certify that this New Application Transmittal and the documents referred to as attached therein are being deposited with the United States Postal Service on this date June 4, 1999 in an envelope as "Express Mail Post Office to Addressee," mailing Label Number EF198925772 US addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231.

Cathy Lewis
(type or print name of person making paper)

Cathy Lewis
Signature of person mailing paper

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"Since the filing of correspondence under § 1.110 without the Express Mail mailing label thereon is an oversight that can be avoided by the exercise of reasonable care, requests for waiver of this requirement will not be granted on petition." Notice of Oct. 24, 1996, 60 Fed. Reg. 55,439, at 55,442.

(Application Transmittal [4-1]—page 1 of 11)

1. Type of Application

This new application is for a(n)

(check one applicable item below)

- ☒ Original (nonprovisional)
☐ Design
☐ Plant

WARNING: Do not use this transmittal for a completion in the U.S. of an International Application under 35 U.S.C. 371(c)(4), unless the International Application is being filed as a divisional, continuation or continuation-in-part application.

WARNING: Do not use this transmittal for the filing of a provisional application.

NOTE: If one of the following 3 items apply, then complete and attach **ADDED PAGES FOR NEW APPLICATION TRANSMITTAL WHERE BENEFIT OF A PRIOR U.S. APPLICATION CLAIMED** and a **NOTIFICATION IN PARENT APPLICATION OF THE FILING OF THIS CONTINUATION APPLICATION**.

- ☐ Divisional.
☐ Continuation.
☐ Continuation-in-part (C-I-P).

2. Benefit of Prior U.S. Application(s) (35 U.S.C. 119(e), 120, or 121)

NOTE: A nonprovisional application may claim an invention disclosed in one or more prior filed copending nonprovisional applications or copending international applications designating the United States of America. In order for a nonprovisional application to claim the benefit of a prior filed copending nonprovisional application or copending international application designating the United States of America, each prior application must name as an inventor at least one inventor named in the later filed nonprovisional application and disclose the named inventor's invention claimed in at least one claim of the later filed nonprovisional application in the manner provided by the first paragraph of 35 U.S.C. 112. Each prior application must also be:

(i) An international application entitled to a filing date in accordance with PCT Article 11 and designating the United States of America; or

(ii) Complete as set forth in § 1.51(b); or

(iii) Entitled to a filing date as set forth in § 1.53(b) or § 1.53(d) and include the basic filing fee set forth in § 1.16; or

(iv) Entitled to a filing date as set forth in § 1.53(b) and have paid therein the processing and retention fee set forth in § 1.21(i) within the time period set forth in § 1.53(f).

37 C.F.R. § 1.78(a)(1).

NOTE: If the new application being transmitted is a divisional, continuation or a continuation-in-part of a parent case, or where the parent case is an International Application which designated the U.S., or benefit of a prior provisional application is claimed, then check the following item and complete and attach **ADDED PAGES FOR NEW APPLICATION TRANSMITTAL WHERE BENEFIT OF PRIOR U.S. APPLICATION(S) CLAIMED**.

WARNING: If an application claims the benefit of the filing date of an earlier filed application under 35 U.S.C. 120, 121 or 365(c), the 20-year term of that application will be based upon the filing date of the earliest U.S. application that the application makes reference to under 35 U.S.C. 120, 121 or 365(c). (35 U.S.C. 154(a)(2) does not take into account, for the determination of the patent term, any application on which priority is claimed under 35 U.S.C. 119, 365(a) or 365(b).) For a o-t-p application, applicant should review whether any claim in the patent that will issue is supported by an earlier application and, if not, the applicant should consider canceling the reference to the earlier filed application. The term of a patent is not based on a claim-by-claim approach. See Notice of April 14, 1995, 60 Fed. Reg. 20,195, at 20,205.

(Application Transmittal [4-1]—page 2 of 11)

WARNING: When the last day of pendency of a provisional application falls on a Saturday, Sunday, or Federal holiday within the District of Columbia, any nonprovisional application claiming benefit of the provisional application must be filed prior to the Saturday, Sunday, or Federal holiday within the District of Columbia. See 37 C.F.R. § 1.78(a)(3).

- ☐ The new application being transmitted claims the benefit of prior U.S. application(s). Enclosed are **ADDED PAGES FOR NEW APPLICATION TRANSMITTAL WHERE BENEFIT OF PRIOR U.S. APPLICATION(S) CLAIMED.**

3. Papers Enclosed

- A. Required for filing date under 37 C.F.R. § 1.53(b) (Regular) or 37 C.F.R. § 1.153 (Design) Application

28 Pages of specification

2 Pages of claims

1 Sheets of drawing

WARNING: DO NOT submit original drawings. A high quality copy of the drawings should be supplied when filing a patent application. The drawings that are submitted to the Office must be on strong, white, smooth, and non-shiny paper and meet the standards according to § 1.84. If corrections to the drawings are necessary, they should be made to the original drawing and a high-quality copy of the corrected original drawing then submitted to the Office. Only one copy is required or desired. For comments on proposed then-new 37 CFR 1.84, see Notice of March 9, 1968 (1990 O.G. 57-62).

NOTE: "Identifying indicia, if provided, should include the application number or the title of the invention, inventor's name, docket number (if any), and the name and telephone number of a person to call if the Office is unable to match the drawings to the proper application. This information should be placed on the back of each sheet of drawing a minimum distance of 1.5 cm. (5/8 inch) down from the top of the page . . . " 37 C.F.R. 1.84(c).

(complete the following, if applicable)

- ☐ The enclosed drawing(s) are photograph(s), and there is also attached a "PETITION TO ACCEPT PHOTOGRAPH(S) AS DRAWING(S)." 37 C.F.R. 1.84(b).
- ☐ formal
- ☒ informal

B. Other Papers Enclosed

____ Pages of declaration and power of attorney

____ Pages of abstract

____ Other

4. Additional papers enclosed

- ☐ Amendment to claims
- ☐ Cancel in this applications claims _____ before calculating the filing fee. (At least one original independent claim must be retained for filing purposes.)
- ☐ Add the claims shown on the attached amendment. (Claims added have been numbered consecutively following the highest numbered original claims.)
- ☐ Preliminary Amendment
- ☐ Information Disclosure Statement (37 C.F.R. 1.98)
- ☐ Form PTO-1449 (PTO/SB/08A and 08B)
- ☐ Citations

- ☐ Declaration of Biological Deposit
- ☐ Submission of "Sequence Listing," computer readable copy and/or amendment pertaining thereto for biotechnology invention containing nucleotide and/or amino acid sequence.
- ☐ Authorization of Attorney(s) to Accept and Follow Instructions from Representative
- ☐ Special Comments
- ☐ Other

5. Declaration or oath (including power of attorney)

NOTE: A newly executed declaration is not required in a continuation or divisional application provided that the prior nonprovisional application contained a declaration as required, the application being filed is by all or fewer than all the inventors named in the prior application, there is no new matter in the application being filed, and a copy of the executed declaration filed in the prior application (showing the signature or an indication thereon that it was signed) is submitted. The copy must be accompanied by a statement requesting deletion of the names of persons who are not inventors of the application being filed. If the declaration in the prior application was filed under § 1.47, then a copy of that declaration must be filed accompanied by a copy of the decision granting § 1.47 status or, if a nonsigning person under § 1.47 has subsequently joined in a prior application, then a copy of the subsequently executed declaration must be filed. See 37 C.F.R. §§ 1.63(c)(1)-(3).

NOTE: A declaration filed to complete an application must be executed, identify the specification to which it is directed, identify each inventor by full name including family name and at least one given name, without abbreviation together with any other given name or initial, and the residence, post office address and country or citizenship of each inventor, and state whether the inventor is a sole or joint inventor. 37 C.F.R. § 1.63(b)(1)-(4).

☐ Enclosed

Executed by

(check all applicable boxes)

- ☐ inventor(s).
- ☐ legal representative of inventor(s).
37 CFR 1.42 or 1.43.
- ☐ joint inventor or person showing a proprietary interest on behalf of inventor who refused to sign or cannot be reached.

☐ This is the petition required by 37 CFR 1.47 and the statement required by 37 CFR 1.47 is also attached. See item 13 below for fee.

☒ Not Enclosed.

NOTE: Where the filing is a completion in the U.S. of an International Application or where the completion of the U.S. application contains subject matter in addition to the International Application, the application may be treated as a continuation or continuation-in-part, as the case may be, utilizing ADDED PAGE FOR NEW APPLICATION TRANSMITTAL WHERE BENEFIT OF PRIOR U.S. APPLICATION CLAIMED.

☐ Application is made by a person authorized under 37 C.F.R. 1.41(c) on behalf of all the above named inventor(s).

(The declaration or oath, along with the surcharge required by 37 CFR 1.16(e) can be filed subsequently).

☐ Showing that the filing is authorized.
(not required unless called into question. 37 CFR 1.41(d))

(Application Transmittal [4-1]—page 4 of 11)

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Certified copy(ies) of application(s)

Country	Appin. No.	Filed
Country	Appin. No.	Filed
Country	Appin. No.	Filed

from which priority is claimed

- ☐ is (are) attached.
- ☐ will follow.

NOTE: The foreign application forming the basis for the claim for priority must be referred to in the oath or declaration. 37 CFR 1.55(a) and 1.63.

NOTE: This item is for any foreign priority for which the application being filed directly relates. If any parent U.S. application or International Application from which this application claims benefit under 35 U.S.C. 120 is itself entitled to priority from a prior foreign application, then complete item 18 on the ADDED PAGES FOR NEW APPLICATION TRANSMITTAL WHERE BENEFIT OF PRIOR U.S. APPLICATION(S) CLAIMED.

10. Fee Calculation (37 C.F.R. 1.16)

A ☒ Regular application

CLAIMS AS FILED				
Claim Number filed	Number Extra	Rate	Basic Fee 37 C.F.R. 1.16(a) \$790.00	
24	4			
Total				
Claims (37 CFR 1.16(c))	24 - 20 = 4	x	\$ 22.00	88.00
Independent				
Claims (37 CFR 1.16(b))	3 - 3 = 0	x	\$ 82.00	0
Multiple dependent claim(s), if any (37 CFR 1.16(d))				
		+	\$270.00	

- ☐ Amendment canceling extra claims is enclosed.
- ☐ Amendment deleting multiple-dependencies is enclosed.
- ☐ Fee for extra claims is not being paid at this time.

NOTE: If the fees for extra claims are not paid on filing they must be paid or the claims cancelled by amendment, prior to the expiration of the time period set for response by the Patent and Trademark Office in any notice of fee deficiency. 37 CFR 1.16(c).

Filing Fee Calculation

§ _____

B. ☐ Design application
(\$330.00—37 CFR 1.16(f))

Filing Fee Calculation

2 _____

C. ☐ Plant application
(\$540.00—37 CFR 1.16(g))

Filing fee calculation

2 _____

(Application Transmittal [4-1]—page 6 of 17)

11. Small Entity Statement(s)

- ☐ Statement(s) that this is a filing by a small entity under 37 CFR 1.9 and 1.27 is (are) attached.

WARNING: "Status as a small entity must be specifically established in each application or patent in which the status is available and desired. Status as a small entity in one application or patent does not affect any other application or patent, including applications or patents which are directly or indirectly dependent upon the application or patent in which the status has been established. The filing of an application under § 1.53 as a continuation, division, or continuation-in-part (including a continued prosecution application under § 1.53(d)), or the filing of a reissue application requires a new determination as to continued entitlement to small entity status for the continuing or reissue application. A nonprovisional application claiming benefit under 35 U.S.C. 119(e), 120, 121, or 365(c) of a prior application, or a reissue application may rely on a statement filed in the prior application or in the patent if the nonprovisional application or the reissue application includes a reference to the statement in the prior application or in the patent or includes a copy of the statement in the prior application or in the patent and status as a small entity is still proper and desired. The payment of the small entity basic statutory filing fee will be treated as such a reference for purposes of this section." 37 C.F.R. § 1.28(a)(2).

(complete the following, if applicable)

- ☐ Status as a small entity was claimed in prior application _____ / _____, filed on _____, from which benefit is being claimed for this application under:

- 35 U.S.C. ☐ 119(e),
☐ 120,
☐ 121,
☐ 365(c),

and which status as a small entity is still proper and desired.

- ☐ A copy of the statement in the prior application is included.

Filing Fee Calculation (50% of A, B or C above)

\$ _____

NOTE: Any excess of the full fee paid will be refunded if small entity status is established and a refund request are filed within 2 months of the date of timely payment of a full fee. The two-month period is not extendable under § 1.136, 37 CFR 1.28(a).

12. Request for International-Type Search (37 C.F.R. 1.104(d))

(complete, if applicable)

- ☐ Please prepare an international-type search report for this application at the time when national examination on the merits takes place.

(Application Transmittal [4-1]—page 7 of 11)

13. Fee Payment Being Made at This Time

☒ Not Enclosed

☒ No filing fee is to be paid at this time.
(This and the surcharge required by 37 C.F.R. 1.16(e) can be paid subsequently.)

☐ Enclosed

☐ Filing fee \$ _____

☐ Recording assignment
(\$40.00; 37 C.F.R. 1.21(h))
(See attached "COVER SHEET FOR
ASSIGNMENT ACCOMPANYING NEW
APPLICATION".) \$ _____

☐ Petition fee for filing by other than all the
inventors or person on behalf of the inventor
where inventor refused to sign or cannot be
reached
(\$130.00; 37 C.F.R. 1.47 and 1.17(f)) \$ _____

☐ For processing an application with a
specification in
a non-English language
(\$130.00; 37 C.F.R. 1.52(d) and 1.17(k)) \$ _____

☐ Processing and retention fee
(\$130.00; 37 C.F.R. 1.53(d) and 1.21(f)) \$ _____

☐ Fee for international-type search report
(\$40.00; 37 C.F.R. 1.21(e)) \$ _____

NOTE: 37 CFR 1.21(f) establishes a fee for processing and retaining any application that is abandoned for failing to complete the application pursuant to 37 CFR 1.53(f) and this, as well as the changes to 37 CFR 1.53 and 1.78(a)(1), indicate that in order to obtain the benefit of a prior U.S. application, either the basic filing fee must be paid, or the processing and retention fee of \$ 1.21(f) must be paid, within 1 year from notification under § 53(f).

Total fees enclosed \$ _____

14. Method of Payment of Fees

☐ Check in the amount of \$ _____

☐ Charge Account No. _____ in the amount of
\$ _____

A duplicate of this transmittal is attached.

NOTE: Fees should be remitted in such a manner that it is clear for which purpose the fees are paid. 37 CFR 1.22(b).

(Application Transmittal [4-1]—page 8 of 11)

(Application Transmittal [4-1]—page 9 of 11)

August 10, 2000

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Ref:TL 287663

HIGH OLEIC/HIGH STEARIC SUNFLOWER OILS

Invented By: Name and citizenship: Enrique Martínez-Force, Spanish.

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Name and citizenship: Rafael Garcés, Spanish.

Address: C/ Dinamarca, 9, E-41907 Valencina de la Concepción, Sevilla, Spain.

0010501000000000

- The present invention relates to transgenic and non-transgenic plants that produce oil having high levels of saturated fatty acids and medium to high levels of oleic acid. More specifically
- 5 the invention relates to a sunflower plant, seed and the extractable oil having increased stearic acid content and increased oleic acid content relative to the normal sunflower oil level of stearic acid and oleic acid.

Background of the Invention

10

Sunflowers (*Helianthus annuus*) are cultivated for the seeds. Seeds have two principal uses. Seeds are used as confectionery seeds but much more importantly for extraction of oil formed within the sunflower seed. In the endogenous oil are a number of fatty acids. The total fatty acid content is given for 3 types of sunflowers: normal, high oleic and high palmitic.

15

These contents are listed below. The saturated or unsaturated fatty acid is given as a percentage of weight relative to the total fatty acid content.

Table 1

FATTY ACIDS	CARBON CHAIN	PERCENT WEIGHT
SATURATED ACIDS		
Palmitic	C16:0	5%-7%= (normal) 2%-4% = (high oleic) 22%-40%= (high palmitic)
Stearic	C18:0	< 10 % but usually 3%- 7%(normal, high oleic, high palmitic)
UNSATURATED ACIDS		
Oleic	C18:1	17%-20%= (normal) 9%-12%=(high palmitic) 75%-90% = (high oleic)
Linoleic	C18:2	50%-70% = (normal, high palmitic) (high oleic)= 2%-10%

20

As table 1 indicates a number of different fatty acid distributions in sunflower oil are known. The table gives the wild type or normal sunflower oil as having a fatty acid distribution of the principle saturates of C16:0 levels of 5%-7%, C18:0 levels of <10%, and of the principle

unsaturates of C18:1 levels of 17%-20% and C18:2 levels of 50%-70%. Presently, high oleic and high palmitic are known sunflower oils with changed saturate, unsaturated fatty acid profiles.

- 5 The industry has been using both transgenic approaches and mutation approaches to alter the saturate, unsaturated fatty acid profiles in the fatty acid biosynthesis pathway. The percentag of saturated and unsaturated fatty acid present in an altered oil reflect the oil's chemical and physical traits. The chemical and physical traits of the oil are altered to form either more useful oil for the industry and/or a healthier oil for the end user.

10

The last twenty years of sunflower research has produced a healthier oil in high oleic sunflower oil. This research has resulted in the commercial availability of sunflowers having high concentrations of oleic acids (C18:1) percentage by weight based in the total fatty acid content of the sunflower oil. Early Russian research into increasing oleic levels in sunflower

- 15 seeds used mutagenesis to form Pervenets sunflowers having elevated oleic acid levels. Such oleic sunflower material is available to the public. Much, if not all, of the high oleic sunflower germplasm available today is the descendants of the early Russian lines resulting from that research. Commercially available high oleic sunflower seed includes for example, sunflower variety, Pioneer Hybrid 6661, which is marketed as producing a seed storage oil
20 having a fatty acid composition that includes 85% oleate.

- The availability of other high oleic sunflower seed is listed with patent documents. For example, there is intellectual property describing high oleic lines which is believed to be based on the Russian Pervenets sunflowers in U.S. Patent No. 4,627,192 and Re-
25 examination certificate B1 4,627,192, issued October 17, 1995, and U.S. Patent No. 4743, 402 and Re-examination certificate B1 4,743,402, issued April 8, 1997 to Fick. This intellectual property lists a number of sunflower varieties that are commercially available for breeding purposes that can be licensed under the Fick patents through a company called A.C. Humko. Additionally, the Fick U.S. patent 4,627,192 indicates that oleic seeds of Sigco
30 41A, 41b, 853R, 4117b, 273W, 416R are available from the Lubrizol Corporation 29400 Lakewood Blvd., Wickliffe, Ohio (USA) 44092.

- In addition to the research, which resulted in increased oleic acid in sunflower oil, there has also been research on increasing the overall industrial usefulness of sunflower oil usually
35 addressed by increasing the level of the saturated fatty acids in oil. Increasing the saturated fatty acids produces an oil that is more suitable for use in the production of margarine, shortening, other food products, and cosmetics by decreasing the need for substantial amounts of hydrogenation.

Some of this type of research is outlined in *Osorio et al.*, in *Crop Sci.* 35: 739-42 (1995). This article describes sunflower seeds developed by traditional breeding and mutagenesis to produce seeds with a high stearate content. This type of research is also outlined in PCT application number EP95/00369 which is entitled "Sunflower seeds and oil having a high stearic acid content". This application teaches, as its name implies, a sunflower oil with increased stearic acid content. One way to obtain this oil is by treating parent seed with a mutagenic agent to induce one or more mutations in the stearic acid biosynthesis pathway. This resulted in an increased production of stearic acid in the sunflower oil in a range between 12% and up to 35% by weight of stearic acid related to the total amount of fatty acid in the oil. High stearic acid producing seeds discussed in this patent are under deposit in the American Type Culture Collection (ATCC). Sunflower seeds identified as "CAS-3" have a average stearic acid content of 25% by weight, related to the total amount of fatty acids in the oil. These seeds were deposited on December 14, 1994 with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852, U.S.A. under deposit accession number ATCC 75968. And sunflower seeds identified as "CAS-4" having an average stearic acid content of 15% by weight, related to the total amount of fatty acids in the oil, were deposited on December 14, 1994 with the American type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852, U.S.A. under deposit accession number ATCC 75969.

This application PCT/EP95/00369 suggests that oil from high stearic lines could be combined with oil from high oleic lines for certain industrial uses. Unfortunately, although this combination of two oils is useful in many instances, there remains a need for a seed that produces high levels of both stearic and oleic fatty acids in the same oil. Particularly since the levels of linoleic acid produced by these stearic acid lines may tend to produce a less desirable profile of fatty acids, then would be produced by a hybrid producing high stearic, high oleic acid.

Traditional breeding and mutagenesis has not been the only tool used to form seeds producing oil with different fatty acid profiles. Increases in stearic acid in oil bearing plants have also been addressed by the introduction of transgenes in to the germplasm, to alter the fatty acid biosynthesis pathway of the vegetable oil. The fatty acid biosynthesis in vegetable oil, but more particularly sunflower oil, includes the biosynthesis of basically two saturates palmitate, stearate and two unsaturates oleate and linoleate. To give a simplified description of the biosynthesis pathway, it is sufficient to say, that palmitate (C16:0) is by enzymatic action chemically modified to form stearate (C18:0), which by enzymatic action is modified to produce oleate (C18:1), that is further modified to form linoleate (C18:2), some minor amounts of araquic (C20:0) and behenic (C22:0) acids are also formed from stearate. In oilseeds the

stearoyl-ACP desaturase is the enzymatic action which introduces the first double bond on stearoyl-ACP to form oleoyl-ACP. Thus, this is an enzyme that assists in the determination of the unsaturation in the C18 length fatty acids.

5 In U.S. patent 5,443,974 the inhibition of canola enzyme stearoyl-ACP desaturase was described. The stearate levels were increased but the levels of palmitate were basically unaffected. Inhibition of the plant enzyme stearoyl-ACP desaturase in canola was also reported by Knutzon *et al.*, *Proc. Nat'l Aca., Sci. USA* 89:2624-28 (1992). These results showed an increase in the level of stearate produced in the canola seed. The research also
10 showed that inhibition by antisense in seeds of canola and soybean, respectively showed increased stearate. When a plasmid containing a gene encoding for stearoyl-ACP desaturase was placed in canola, this inhibition resulted in both an increase in stearic acid and unfortunately a reduction in the oleate. However, in the soybean this inhibition of stearate resulted in a less dramatic reduction of the oleate. This slower decrease in oleate
15 however may have been a function of the small initial levels of oleate in the soybean. The fatty acid pathway in most oilseed plants appears to be resistant to maintaining both oleic and stearic at elevated levels.

Additionally, Pioneer Hi-Bred International, Inc. have increased the levels of both stearic acid
20 and palmitic acid in sunflowers was through the inhibition of the plant enzyme stearoyl-ACP desaturase. This research was surprising in light of other transformations in other plants that indicated that this enzyme would not effect palmitic acid levels only stearic acid levels. Unfortunately, palmitic oil is not viewed as being a very healthy oil. This research is indicated in PCT/US97/01419.

25 Pioneer Hi-Bred International, Inc. taught in PCT/US96/09486 that sunflower oil levels of both palmitic and oleic acids could be increased. Sunflower seeds having increased levels of palmitic 21-23% and oleic 61% were deposited in the ATCC as 93PMOL040G and 93PMOL040F in the American Type Culture Collection 12301 Parklawn Drive, Rockville
30 Maryland 20852, U.S.A. having Accession No. 97159 and 97158 respectively. The patent application teaches a sunflower oil that is liquid at room temperature. But the increased palmitic fatty acid level in the Pioneer invention, is alleged to allow the oil to be used in shortening and in margarine with relatively low level of hydrogenation, which leads to a relatively low level of trans-fatty acids in the resulting product. However the commercial value
35 may be questioned as palmitic oil is not viewed as being a very healthy oil. Thus margarine made with this oil may be substantially less desirable than margarine made with a healthier combination of fatty acids (i.e. stearic acid).

BRIEF DESCRIPTION OF THE FIGURES

Figure 1-TAG shown hydrolysis of the triacylglycerols by the lipase.

Figure 2-is elevated thioesterase shown as the relative affinity of the thioesterase on

5 stearyl-ACP and oleoyl-ACP of HOHT and HOLT.

Figure 3- plastid showing the fatty acid biosynthesis in oilseeds

Figure 4- the F2 segregation for HOHT indicating HOHT progeny

Figure 5- the F2 segregation for HOLT indicating no HOHT progeny

10 DEFINITIONS:

SUNFLOWER plant shall mean all *Helianthus annuus* plant and cell parts including pollen, kernel, oil, embryo, stalk, head, roots, cells, meristems, ovule, anthers, microspores, embryos, DNA, RNA, petals, seeds, and the like and protoplasts, callus or suspensions of
15 any of the above.

15DAF shall mean 15 days after flowering.

HOLT shall mean a sunflower having high to medium-high (40%-90%) oleic acid levels in the
20 oil when compared to normal, wildtype sunflower seed (oleic acid levels of 17%-20%) levels wherein there is low levels of thioesterase activity.

TOTAL FATTY ACID CONTENT refers to the sum of C16:0,18:0,18:1,18:2, 20:0, 22:0 and the traces of other like fatty acids as determined simultaneously in the seed sample.
25

HOHT shall mean a sunflower having high to medium-high (40%-90%) oleic acid levels in the oil when compared to normal, wildtype sunflower seed (oleic acid levels of 17%-20%) levels wherein there is high levels of thioesterase activity.

30 LOW LEVELS OF THIOESTERASE ACTIVITY shall mean levels of stearyl-ACP thioesterase activity at DAF15 which are lower than 10% of the activity over oleoyl-ACP, this permits an exportation out of the plastid of a maximum of less than 12% of stearic acid with respect to the total fatty acids synthesised by the plastid.

35 HIGH LEVELS OF THIOESTERASE ACTIVITY shall mean levels of stearyl-ACP thioesterase activity at DAF15 which are higher than 10% of the activity over oleoyl-ACP, this permits an exportation out of the plastid of a minimum of 12% of stearic acid with respect to the total fatty acids synthesised by the plastid.

Summary of the Invention

5 The present invention broadly relates to high oleic/high stearic vegetable oils. More specifically the present invention relates to the sunflower seeds, the sunflower plants, sunflower meal or crushed seeds, oil obtainable from the seeds and the extracted endogenous sunflower oil, which has a fatty acid profile high in oleic acid and in stearic acid. Still more specifically this invention has a fatty acid profile that also has preferably less than 25 percent linoleic and more preferably less than 10 percent linoleic. Additionally the profile
10 of the invention is often selected to have palmitic levels below 21%.

15 In the broad scope the invention the sunflower seeds, the sunflower plants, sunflower meal or crushed seeds obtainable from the seeds and the extracted sunflower oil contain or produce an oleic acid content of more than 40% weight and a stearic acid content of more than 12% weight based on the total fatty acid content of the oil. The more preferred invention for certain spread uses has oil and the seeds containing the oil which contain an oleic acid content that is from 55 to 75% weight relative to total fatty acid content and a stearic acid content from 15 to 40% weight relative to total fatty acid content. The scope of the invention includes oil and the seeds containing the oil plants capable of forming such oils which have a
20 total level of saturated fatty acids of at least 20 wt%.

25 The sunflower seed containing oil is usually selected to have at least 90 % weight of the saturated fatty acid groups in the 1 and/or 3 position as indicated in Table 6. In some instances the invention can have from 95 to 100% weight of the saturated fatty acid groups in the 1 and/or 3 position. The present invention also includes a sunflower plant capable of producing the sunflower seeds containing such oils.

30 The present invention also covers the method of obtaining a sunflower oil having an oleic acid content of more than 40% weight and a stearic acid content of more than 12% weight based on the total fatty acid content of the oil by extracting oil from the seeds. The method includes an extraction process which does not involve a substantial modification of the sunflower oil. Additionally, in the process of extraction of the oil from the seed there is no substantial chemical or physical modification nor enzymatic rearrangement taking place and no substantial hardening of the oil.

35 The present invention also includes food products comprising oil obtainable from seeds having an oleic acid content of more than 40% weight and a stearic acid content of more than 12% weight based on the total fatty acid content of the oil. Food products that are particularly

useful for this type of oil include spreads, margarine's, shortenings, sauces, ice-cream, soups, bakery products, confectionery products, and the like. In these food products the level of sunflower oil is from 3 to 100% weight relative to the total oil weight in the product. When used to form a spread according to the present invention the sunflower oil is used as a
5 hardstock at levels of 5 to 20% by weight.

The present invention also encompasses cosmetic products comprising a sunflower oil, the oil having an oleic acid content of more than 40% weight and a stearic acid content of more than 12% weight based on the total fatty acid content of the oil. These cosmetic products
10 can contain levels of sunflower oil from 3 to 100% by weight. Some examples of these cosmetic products would include creams, lotions, lipsticks, soap bars and skin or hair oils.

The present invention also includes a process for selecting *Helianthus annuus* plants, capable of producing the seeds having the desired oil. The steps of the method are a)
15 selecting a number of *Helianthus annuus* plants, collecting therefrom the seeds, which have a mutation induced by a mutagenic agent or an introduced transgene such that the oil in the seed has a stearic acid content of at least 12% weight based on the oil present in the seed;
(b) selecting a number of *Helianthus annuus* plants, collecting therefrom the seeds, which express an oleic acid content of at least 40% weight based on the oil present in the seed and
20 a high level of thioesterase activity (c) crossing the plants grown from the seeds of (a) and (b) and, harvesting the F1 seed progeny.

Additional steps include the steps of: (d) planting of the seeds or embryos rescue of the embryos of the F1 progeny obtained to form F2 segregating seeds; (e) selecting from the F2 seeds which developed plants, those plants which produce seeds having an oleic acid
25 content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of the oil, optionally selfing the selected plant to form true breeding inbreds.

The present invention also includes the process for producing F1 hybrid seed. The steps of
30 the method are a) planting seed of two inbreds having high oleic acid content of at least 40% and high levels of thioesterase activity, one of which may be male sterile, b) crossing the two inbreds, and c) harvesting the F1 seed capable of producing F2 seed with a 40% oleic acid content and a 12% stearic acid content.

35 Description of the Invention

The present invention encompasses a vegetable oil with a new and unique fatty acid composition produced in easy to grow crops. The preferred crop is sunflower. However,

mutation and/or transformation to produce the derived vegetable oil could likewise modify other crops. These crops would include at least *Brassicas*, peanuts, palms and other oil producing plants. The crop should be readily transformable and/or susceptible to mutagenically induced oil changes. Rape seed meets all these requirements as does sunflower, these crops are presently some of the most useful crops for production of this new and unique fatty acid composition in the oil of their seeds..

The industrial use for oil which are capable of being extracted from the seeds is determined by its fatty acid composition. Table 2 shows the fatty acid composition and other properties of some fats and oils. The oils obtained from seeds cultivated in temperate climate (sunflower, soybean, rape seed, etc.) have mainly unsaturated fatty acids, like linoleic and oleic acids, so they are liquid and primarily used for cooking, salad dressing, etc. Saturated fatty acids are solid at room temperature and referred to as fats. These saturated fatty acid are mainly obtained from animals (margarine, lard, etc.), some tropical trees (cacao, palm) or one chemically modified (hydrogenation and transesterification) liquid vegetable oils. These modified liquid vegetable oils have mainly saturated (palmitic or stearic acids) or chemically modified fatty acids (trans fatty acids) with high melting points.

Table 2

Oil or fat	Fatty acid composition (%)						Properties	
	Others	Myristic	Palmitic	Stearic	Oleic	Linoleic	Trans	Saturated
Lard	3	2	25	12	45	10	1	79
Butter	14	10	26	12	28	3	3	84
Margarine			10	7	46	34	23	
Palm Oil		1	45	5	39	9		18
Olive Oil	1		14	3	71	10		2
Cocoa Butter			26	35	35	3		4
Normal Sunflower			7	5	30	57		1
High oleic sunflower			5	4	88	2		1

Table 2. Total fatty acid, and other special properties, like trans fatty acid and saturated fatty acid at position 2 of the TAG molecule, of several oils and fats. In the column labelled others are palmitoleic acid in the case of lard and olive oil and also fatty acids shorter than 12 carbons in the butter. Minor amounts (<2%) of other fatty acids are also present in some oils.

The fats are needed by the food industry to make margarine, shortening, bakery, confectionery, snacks, etc. The industry uses the fat for their physical properties. Fats do not readily melt at room temperature, can be spread and does not stick. Fats also have greater stability, which is a resistance to oxidation at room temperatures.

Unfortunately, fats also have negative nutritional properties. Many fats raise the levels of bad serum cholesterol (low-density lipoprotein, LDL) in blood. The principal component of fats and oils is triacylglycerol (TAG), normally making up more than 95 % of the fat or oil. Three fatty acids are bound to a molecule of glycerol to make the TAG. These fatty acids are either saturated or unsaturated. Fat having negative nutritional properties is due to having most of the saturated fatty acids in the position 2 of the TAG molecule (figure 1). Most vegetable fats and oils which do not raise LDL have little saturated fatty acids in this position. The fatty acids in positions 1 and 3 are liberated as free fatty acids by lipase in digestion, if these fatty acids are saturated they form insoluble salts with calcium and magnesium, which are mainly excreted. But fatty acids in position 2 form a molecule of monoacylglycerol, which is easily absorbed into the body, raising LDL.

Therefore, vegetable oils are modified to form hard fats. These vegetable oils are hydrogenated and/or transesterified to increase the percentage of saturated fatty acids. However, the resulting oils are unfortunately not necessarily healthier fats. The hydrogenation process produces "trans fatty" acids that probably are even less healthy than saturated fatty acids. While the transesterification process randomly changes the fatty acid within the three positions. Thus, neither of the two chemical modifications works well to truly form healthy fats.

But not all saturated fats are unhealthy, it have been demonstrated that cocoa butter, that has around 60% of saturated fatty acids, the rest being mainly oleic acid, does not raise serum cholesterol. This is due to two main reasons, one is that only 4% of the saturated fatty acids are in position 2 and the other is the principal saturated fatty acid is stearic acid. Stearic acid does not appear to have a negative effect on serum cholesterol, and is considered healthier than palmitic acid, which is the main saturated fatty acid of commodity

fats. Additionally, the 35% of oleic acid content in the cocoa butter also provides healthy properties to the fat.

5 There remains a need of the industry for a vegetable oil (fat) with high stearic acid (as the saturated fatty acid) and elevated oleic acid (as unsaturated fatty acid) content that will resolve the actual fat problems, preferably with the stearic acid in the 1 and 3 position of TAG. T that end the present invention can be selected to have at least 90 w/w % of saturated fatty acids in the 1 and/or 3 position. In the more preferred embodiments of the invention there is 95-100% w/w% of saturated fatty acids in the 1 and/or 3 position. Thus the
10 present invention produces an oil/fat that is addressing the industry's need for healthier fats.

The main characteristics of modified plants, its seeds and the oil capable of being extracted from these seeds according to the invention are the production of a vegetable oil with increased oleic acid (beyond normal Sunflowers in Table 1) and with a stearic acid content of
15 12% and above. Even more specifically the percentage stearic acid content is between 15% and 40% by weight and even more specifically the invention can be selected to have a stearic acid content of between 20% and 35%, an oleic acid content of preferably above 40%. More specifically an oleic acid content of the oil of the present invention is between 50% and 80% by weight. Yet, more specifically the present invention has the high stearic
20 acid and the high oleic acid content of between 55% and 75% by weight relative to the total fatty acid content. Additionally, the invention with high oleic and high stearic acid also contains high levels of thioesterase activity. The linoleic acid content of the present invention is usually selected to be less than 25% by weight related to the total amount of fatty acids in the oil. The seeds according to the invention can be selected to have at least 20% by weight
25 total level of saturated fatty acids relative to the total fatty acid content for the oil.

Originally it was believed that crossing the prior art high oleic lines with the prior art high stearic lines would produce an inbred line having high oleic and high stearic properties. Unfortunately, this has proved extremely difficult. The prior art high stearic acid content lines,
30 which were selected after mutagenesis, like CAS-3, evidenced low oleic content. Likewise, most high oleic varieties evidenced low stearic acid content.

We have discovered that the selection process for very high oleic material, particularly in sunflower, selects against stearic acid generally. Thus the present invention is possible due
35 to the discovery that to form a plant such as a sunflower plant, which is capable of producing the desired oil, the starting material should contain a third characteristic beyond high oleic and high stearic, it should evidence elevated thioesterase activity over the stearyl-ACP as indicated in Figure 2.

To assist in the understanding of the role of thioesterase activity within the present invention, a general review of the fatty acid biosynthesis in sunflower is helpful. The seed fatty acid biosynthesis occurs inside the plastid (Figure 3). The fatty acids within the plastid remain covalently bound to the acyl carrier protein (ACP) during successive cycles of extension (synthesis of palmitic and stearic acids) and desaturation (conversion of stearic acid to oleic acid). The major products of this biosynthetic process are palmitoyl-ACP, stearoyl-ACP and oleoyl-ACP. A series of cycling reactions catalysed by the enzymatic complex FAS I produce the palmitoyl-ACP that have 16 carbons. A second enzymatic complex called FAS II elongates the palmitoyl- ACP to stearoyl-ACP (18 carbons), that is further modify by the stearate desaturase to produce oleoyl-ACP. These are the three main fatty acids synthesised by the plastid. These are separated from the ACP by the action of the thioesterase (Acyl-ACP thioesterase catalyse the hydrolysis of acyl thioesters) and then exported out of the plastid in to the cytoplasm where the synthesis of triacylglycerides takes place. Later the oleic acid can be desaturated to linoleic and linolenic acids. The synthesis of TAG (storage oil) is produced in the cytoplasm using the pool of fatty acids in the cytoplasm, and this pool is made from the fatty acids exported from the plastid and the linoleic acid made by desaturation. So, the fatty acid composition of TAG is determined by the plastid exported fatty acids plus the linoleic acid produced in the cytoplasm.

The present invention, a vegetable oil that is rich in stearic and oleic acids, is formed by altering the standard fatty acid biosynthesis described above. By combining a reduced stearate desaturase activity with a good thioesterase activity on stearoyl-ACP in a high oleic line a vegetable oil rich in stearic and oleic acid is formed according to this invention. The invention accumulates stearoyl-ACP inside the plastid, and the thioesterase over stearoyl-ACP activity exports it out of the plastid. This higher stearic acid content is available for TAG biosynthesis. Later, in the cytoplasm the high oleic character avoids the conversion of oleic acid into linoleic acid.

A plant capable of producing the oil of the present invention may be obtained by two basic methods. The first method is mutagenesis and breeding the second method is transformation and breeding which is discussed later. Regardless of the method, the trait high levels of thioesterase activity should be identified and bred for in the scope of the present invention.

To form the oil of the present invention a plant capable of producing seed having oil containing the high oleic and high stearic acid content must be formed. To achieve this objective a high oleic, high thioesterase line can be crossed with a high stearic line which results after selfing and selection in an inbred that is high oleic and high stearic. This

requires two basic elements a high stearic line and a HOHT line. The simplest way to form the present invention is by making sexual crosses of a high stearic line such as CAS-3, with a HOHT sunflower line and selecting for HSHO (a line which produces a seed characterised by high stearic acid and high to medium high oleic acid in the oil). The first basic element

5 CAS-3 is material deposited in the ATCC, the second element is the HOHT.

The first basic element can also be formed by mutation of existing germplasm and screening for the high stearic trait. This can be done according to the teaching of application PCT/EP95/00369. Various mutation methods are detailed in the examples. But briefly one

10 method to form a high stearic acid line treats the parent seeds for 2 hours at room temperature in a solution of 70 mM ethyl methane sulfonate in water, or 2 hours at room temperature with a solution of 2 mM sodium azide in water to form a desired oil trait mutation. The resultant material is selected for the stearic acid trait. The lines are selfed (if necessary) to form a true breeding line.

15 The HOHT line can be screened and selected from the existing prior art high oleic sunflower lines. The screening would be based on maintaining the high oleic trait and locating the trait for high thioesterase activity and selecting for both traits. Alternatively the second element, HOHT, can be formed by crossing a fixed high oleic line which is HOLT with a line

20 evidencing high levels of thioesterase activity and through selfing and selection throughout several generations, a new HOHT can be formed. A less efficient alternative to forming the HOHT is by mutation, selfing, screening and selection throughout several generations. In this alternative using normal germplasm and mutating it and selecting for either the high oleic trait or the HOHT trait can form the HOHT line. However, since there is a number of high

25 oleic sunflower lines available it should not be necessary to reform a HOHT line from normal germplasm. Screening high oleic lines for high levels of thioesterase activity should be sufficient to locate a HOHT line.

The reason a HOHT line is necessary and the high oleic trait does not appear sufficient is

30 that it have been found that the selection process for very high oleic material selects against stearic generally and also against elevated thioesterase activity over the stearyl-ACP. Although some high, or more accurately medium-high, oleic lines carry elevated thioesterase activity over the stearyl-ACP, in general, as the oleic level increases the thioesterase activity decreases. Measuring the thioesterase activity, from lines, having high eighties, and

35 oleic lines, with medium-high levels at sixty, seventy and low eighties of oleic, shows a number of high oleic lines in the medium-high level with good activity over stearyl-ACP.

Turning to the thioesterase activity over stearyl-ACP, this activity is used to select the material for use in the development of new inbred line carrying high oleic and stearic traits according to the present invention. During sunflower seed formation, an active period of fatty acid biosynthesis takes place between 12 and 28 DAF. The maximum in vitro acyl-ACP (acyl-carrier-protein) thioesterase activities are found at 15 DAF, preceding the largest accumulation of lipid in the seed (18 DAF). Changes in acyl-ACP thioesterase activity are not only quantitative, but also qualitative, since, although the preferred substrate is always oleoyl-ACP, the affinity for palmitoyl-ACP decreases, whereas that for stearyl-ACP increases with seed maturation. At 15 DAF the level of thioesterase activity can be measured as shown in the examples to determine which lines should be progressed.

In Table 3 we can see an example of two high oleic lines, the usual high oleic line which has low thioesterase over stearyl-ACP (HOLT) and the high oleic line with high thioesterase over stearyl-ACP (HOHT) which has more than twice the thioesterase Vmax over stearyl-ACP than the usual high oleic line.

Table 3

Sunflower line	Stearic acid (%)	V max
HOHT	17.8	2.03
HOLT	8.0	0.82

Table 3. Stearic acid content and thioesterase Vmax over the stearyl-ACP at 15 days after flowering seeds from two high oleic sunflower lines.

This HOHT line is deposited with the ATCC as _____. As Table 3 indicates, this line has more stearic acid at 15 days after flowering than the HOLT. This stearic acid level at 15 DAF can also be employed as a rough screening protocol for selecting HOHT lines. The relative affinity of the enzymes over the stearyl-ACP standardised with respect to the one over oleoyl-ACP are also shown in Figure 2.

Additionally, the oil obtained from the mature seed of the HOHT also evidences increased stearic acid levels relative to the HOLT line (Table 4).

Table 4

Sunflower line	Fatty acid composition (%)					
	Palmitic	Stearic	Oleic	Linoleic	Araquic	Beh nic
HOHT	4.3	9.7	78.5	3.9	1.0	2.6
HOLT	3.8	4.9	84.3	4.8	0.5	1.7

Table 4. Fatty acid composition (%) of seeds from two high oleic sunflower lines.

5

Commercially and publicly available high oleic material, the deposited material or newly formed mutated high oleic lines produced according to the examples or by other methods of mutation or transformation can be screened according to the procedure outlined in example 6. The selected lines can have an increased thioesterase Vmax on the substrate stearoyl-

10

ACP at 15 DAF having activity levels as indicated in the definition of high levels of thioesterase. The result of finding a HOHT line is that the fatty acid content will most often evidence an increase in the stearic acid level (when compared to a HOLT) in the line while maintaining a high oleic content. It should be noted that the oleic content of these (HOHT) lines is more likely to be in the sixty, seventy or low eighty range versus the high eighties.

15

Previous to our invention, selecting for high oleic lines selected against this thioesterase activity. Thus when a HOLT (high oleic line with low thioesterase activity) was crossed with or transformed with a high stearic producing trait the stearic level dropped to unacceptably low levels. However, the HOHT line when crossed with the stearic acid line carries the stearic acid level of the CAS 3 in the F2 seed. In contrast the HOLT line decreased the stearic acid level of the CAS 3 in the F2 seed. This is shown in Table 5 where lines of both HOLT and HOHT have been crossed with the high stearic CAS-3 line. Also shown in Table 5 is the fatty acid composition of these lines.

25

Table 5

Sunflower line	Fatty acid composition (%)					
	Palmitic	Stearic	Oleic	Linoleic	Araquic	Behenic
HOHT x CAS-3	5.2	24.6	59.2	6.8	1.8	2.4
HOLT x CAS-3	4.3	17.4	72.1	4.0	1.3	2.8

Table 5. Fatty acid composition of selected high oleic and stearic lines, with high and low thioesterase activity over stearoyl-ACP, after crossing by CAS-3.

30

In Figures 4 and 5, the F2 segregation for both high stearic acid content and high oleic acid content of the above lines are shown. The seeds formed from the HOHT x CAS-3 cross which have the higher stearic and oleic acids are within a circle. The line with high thioesterase activity over stearyl-ACP has these kinds of seeds and the line without this high thioesterase activity have no seeds of this type.

The selected F2 lines are selfed for 5 to 6 generations in isolated conditions to avoid contamination. The resultant generations are selected, based on high oleic, and stearic acid content. Thioesterase activity can be analysed to assist in the selection process. Likewise, marker assisted breeding can be employed to track any or all of the three traits to make the selection process quicker. Various markers such as SSR microsatellite, ASO, RFLP and likewise can be employed. The use of markers is not necessary, as standard tests are known for determining oleic, stearic, and thioesterase activity. However, once identified markers make trait tracking easier and earlier in the plant's life.

The true breeding plants produce an oil having a similar fatty acid composition that the F2 seeds selected with a low content of saturated fatty acid in the 2 position of the TAG molecule (Table 6).

Table 6

	Fatty acid composition (mol%)					
	Palmitic	Stearic	Oleic	Lionoleic	Araquic	Behenic
Total oil	5.5	24.9	57.8	8.2	1.7	1.8
TAG	5.6	26.1	57.6	7.4	1.6	1.7
sn-2 position	1.7	1.9	87.4	9.0	n.d.	n.d.
sn-1 and 3 position	7.2	33.1	46.8	7.3	2.7	2.9

Table 6. Fatty acid composition of oil, TAG and Sn positions of true breeding HSHO plants selected.

n.d.= not detected.

As indicated earlier the two basic elements employed to form the present invention can be developed by mutation and breeding or by transformation and breeding. In accordance with the present invention the sunflower plant can evidence the stearic or oleic acid traits by introduction, either presently or in the ancestry of the line, mutation of the genes. A number of mutation methods including irradiation, chemical mutation, and other forms of causing gene deletions or changes can work. Particularly, easy to use are mutagenic agents that result in small changes in the genome such as point mutations.

Alternatively, as mentioned previously there is a second method to produce the stearic or oleic lines in accordance with the present invention, the sunflower plant can evidence the stearic or oleic acid traits by transforming a gene into the plant to produce one or more of the desired traits.

The use of gene sequences to inhibit or promote gene expression is quite well understood. A complete gene sequence, under the control of a promoter that operates effectively in the plant, will generally overexpress the gene product, leading to an amplification of the effect of the protein so produced. Sometimes the gene product is reduced: this phenomenon is termed "co-suppression". Reduction of the gene product is also generally obtained by using a dominant negative mutation, or by reversing the orientation of the gene sequence with respect to the promoter so that it produces "antisense" messenger RNA.

A DNA construct according to the invention may be an "antisense" construct generating "antisense" RNA or a "sense" construct (encoding at least part of the functional protein) generating "sense" RNA. "Antisense RNA" is an RNA sequence which is complementary to a sequence of bases in the corresponding mRNA: complementary in the sense that each base (or the majority of bases) in the antisense sequence (read in the 3' to 5' sense) is capable of pairing with the corresponding base (G with C, A with U) in the mRNA sequence read in the 5' to 3' sense. Such antisense RNA may be produced in the cell by transformation with an appropriate DNA construct arranged to generate a transcript with at least part of its sequence complementary to at least part of the coding strand of the relevant gene (or of a DNA sequence showing substantial homology therewith). "Sense RNA" is an RNA sequence, which is substantially homologous to at least part of the corresponding mRNA sequence. Such sense RNA may be produced in the cell by transformation with an appropriate DNA construct arranged in the normal orientation so as to generate a transcript with a sequence identical to at least part of the coding strand of the relevant gene (or of a DNA sequence showing substantial homology therewith). Suitable sense constructs may be used to inhibit gene expression (as described in International Patent Publication WO91/08299) or a sense construct encoding and expressing the functional protein may be transformed into the plant to over express the protein.

DNA constructs according to the invention may comprise a base sequence at least 10 bases (preferably at least 35 bases) in length for transcription into RNA. There is no theoretical upper limit to the base sequence it may be as long as the relevant mRNA produced by the cell but for convenience it will generally be found suitable to use sequences between 100

and 1000 bases in length. The preparation of such constructs is described in more detail below.

As a source of the DNA base sequence for transcription, a suitable cDNA or genomic DNA or synthetic polynucleotide may be used. The isolation of suitable sequences encoding stearyl-ACP desaturase from *Brassica* and *Arabidopsis* are described in U.S. patent 5,443,974: similar methods may be used to isolate sequences encoding stearyl-ACP desaturase from other plants. These may have greater or lesser degrees of homology with sequences encoding stearyl-ACP desaturase from *Arabidopsis*. Sequences coding for the whole, or substantially the whole, of the protein may thus be obtained. Suitable lengths of this DNA sequences may be cut out for use by means of restriction enzymes. When using genomic DNA as the source of a partial base sequence for transcription it is possible to use either intron or exon regions or a combination of both.

To obtain constructs suitable for modifying expression of these desaturase in plant cells, the cDNA sequence as found in the protein cDNA or the gene sequence as found in the chromosome of the plant may be used. Recombinant DNA constructs may be made using standard techniques. For example, the DNA sequence for transcription may be obtained by treating a vector containing said sequence with restriction enzymes to cut out the appropriate segment. The DNA sequence for transcription may also be generated by annealing and ligating synthetic oligonucleotides or by using synthetic oligonucleotides in a polymerase chain reaction (PCR) to give suitable restriction sites at each end. The DNA sequence is then cloned into a vector containing upstream promoter and downstream terminator sequences. If antisense DNA is required, the cloning is carried out so that the cut DNA sequence is inverted with respect to its orientation in the strand from which it was cut.

In a construct expressing antisense RNA, the strand that was formerly the template strand becomes the coding strand, and vice versa. The construct will thus encode RNA in a base sequence which is complementary to part or all of the sequence of the protein mRNA. Thus the two RNA strands are complementary not only in their base sequence but also in their orientations (5' to 3').

In a construct expressing sense RNA, the template and coding strands retain the assignments and orientations of the original plant gene. Constructs expressing sense RNA encode RNA with a base sequence, which is homologous to part, or all of the sequence of the mRNA. In constructs, which express the functional protein, the whole of the coding region of the gene is linked to transcriptional control sequences capable of expression in plants.

- For example, constructs according to the present invention may be made as follows. A suitable vector containing the desired base sequence for transcription is treated with restriction enzymes to cut the sequence out. The DNA strand so obtained is cloned (if desired, in reverse orientation) into a second vector containing the desired promoter sequence and the desired terminator sequence. Suitable promoters include the Actin2 promoter or the pea plastocyanin promoter sequence. Suitable terminator sequences include that of the *Agrobacterium tumefaciens* nopaline synthase gene (the nos 3' end).
- 10 In a DNA construct according to the invention, the transcriptional initiation region may be derived from any plant operative promoter. The transcriptional initiation region may be positioned for transcription of a DNA sequence encoding RNA, which is complementary to a substantial run of bases in a mRNA encoding the desaturase enzyme (making the DNA construct a full or partial antisense construct).
- 15 The transcriptional initiation region (or promoter) operative in plants may be a constitutive promoter or an inducible or developmentally regulated promoter, as circumstances require. For example, it may be desirable to modify protein activity at certain stages of the plant's development. Use of a constitutive promoter will tend to affect protein levels and functions in all parts of the plant, while use of a tissue-specific promoter allows more selective control of gene expression and affected functions. Thus the antisense or sense RNA is only produced in the organ in which its action is required.
- 20 The DNA constructs of the invention may be inserted into plants to regulate the expression of the gene encoding for a desaturase or thioesterase or other enzymes in the fatty acid pathway resulting in modification of plant characteristics for production of stearic acid or oleic acid or thioesterase activity in the fatty acid synthesis pathway. Depending on the nature of the construct, the production of the gene product may be increased, or reduced, either throughout or at particular stages in the life of the plant. Generally, as would be expected, production of the protein is enhanced only by constructs, which express RNA homologous to the substantially complete endogenous protein mRNAs. Full-length sense constructs may also inhibit protein expression. Constructs containing an incomplete DNA sequence shorter than that corresponding to the complete gene generally inhibit the expression of the gene and production of the proteins, whether they are arranged to express sense or antisense RNA.
- 25 30 35

A DNA construct of the invention is transformed into a target plant cell. The target plant cell may be part of a whole plant or may be an isolated cell or part of a tissue, which may be

regenerated into a whole plant. The target plant cell may be selected from any monocotyledonous or dicotyledonous plant species. The preferred target plant is an oil producing species of monocot or dicot. The more preferred target plant is a *Brassica* or *Helianthus annuus* L. Plants may be derived from the transformed plant cell by regeneration of transformants and by production of successive generations of the transformants' progeny.

Constructs according to the invention may be used to transform any plant using any suitable transformation technique to make plants according to the invention. Both monocotyledonous and dicotyledonous plant cells may be transformed in various ways known to the art. In many cases such plant cells (particularly when they are cells of dicotyledonous plants) may be cultured to regenerate whole plants which subsequently reproduce to give successive generations of genetically modified plants. Any suitable method of plant transformation may be used. For example, many dicotyledonous plants such as sunflower, oilseed rape, tomato and melon may be transformed by *Agrobacterium* Ti plasmid technology, such as described by Bevan (1984, *Nucleic Acid Research*, 12:8711-8721) or Fillatti et al (*Biotechnology*, July 1987, 5:726-730). Such transformed plants may be reproduced sexually, or by cell or tissue culture. Other methods use the gene gun, microinjection, PEG and electroporation.

Examples of plants which could be modified to produce the desired oil according to the present invention include the oil from oilseed rape (canola), palm, sunflower, soya. Other possible seed transformants are cereals, for example rice and maize, wheat, barley, oats and rye.

Thus crossing mutants or transgenics or a combination to form seeds containing at least 40% oleic oil and at least 12% stearic oil according to the invention can produce tailor-made seeds and tailor-made oil in accordance with the present invention. The result HSHO lines can be used in further conventional plant improvement practices by crossing with other known or as yet unknown mutant, transgenic or wild type plants.

Alternatively, forming a high stearic line which has a transgene (for example as shown in 5,443,974) that inhibits stearoyl-ACP desaturase activity can make one of the basic elements of the present invention. The gene can be extracted from sunflower material by using the known *Arabidopsis* or the *Brassica* stearoyl-ACP desaturase-encoding gene as taught in U.S. 5,443,974 the sequence of which is in the patent and is incorporated by reference. The known method of transforming sunflower is taught in International application number US97/01419. This application, unfortunately, does not teach a gene; it does teach how to transform the gene once isolated with a plasmid into the plant genome. The teaching of both of these patents together places the transformation of a desaturase gene into a dicot within

the ordinary skilled person's ability. Any of these genes or other similar genes having substantial homology can be placed in either the high oleic line or in a HOHT line that is crossed thereto.

- 5 The following differences to the standard procedure used to form a transgenic sunflower are herein noted. The preferred method of transformation is the Agrobacterium method of transformation. However, due to gene silencing concerns when sunflower is transformed, the promoter should be either an Arabidopsis actin gene promoter "Actin2", available from the University of Georgia, Life science Building, Athens Georgia 30602-7223 or possibly a pea
- 10 plastocyanin promoter which is published in the LAST & Gray 1989 (Plant Mol. Biol. 12, 655-666) and in Plant Journal (1993) 3(3):437-49. by K. Pwee and J. Gray supplied by the University of Cambridge Dept. of Plant Science Downing Street Cambridge CB23EA or the like. These promoters are more useful in sunflower transformation than 35S or 19s CaMV promoters, though the pea plastocyanin promoter may still show some difficulties.

- 15 Thus a high oleic line with elevated thioesterase activity can be transformed with a gene that produces high stearic acid. Although the thioesterase trait is most often discussed in conjunction with the high oleic line it could also be in the high stearic line that is crossed to the high oleic line to form the HSHO line of the present invention. Alternatively, two lines can
- 20 be transformed, one to have oleic and one to have higher stearic and they can be crossed to form an inbred which is selected for elevated thioesterase activity and both other traits. Or one line can be transformed with both a high oleic and high stearic gene and selected for based on elevated thioesterase activity.

- 25 Alternatively, to avoid transgenic issues a line can be mutated to have both oleic and high stearic with the elevated thioesterase activity in a two step mutation process. Two separate lines can be mutated to carry the oleic and thioesterase trait and the stearic trait and crossed and selfed to form a plant which produces the oil. This would most likely require two inbreds both carrying oleic and stearic oil traits to assure the hybrid formed by crossing these two
- 30 inbreds had these traits. Once a trait is fixed, through standard breeding or through a marker assisted breeding this trait can readily be moved into different inbreds. By using at least one trait that is transgenic the breeding becomes simpler as transgenes are usually dominant in their inheritance. Thus only one line in a hybrid cross would have to carry the transgene, however, both lines would preferably carry the mutated traits as these are most often
- 35 recessive or additive traits not dominant traits.

The seed generated when producing the sunflower capable of producing the oil of the present invention is analysed with a methodology described in Garcés, R. and Mancha, M.

"One-step lipid extraction and fatty acid methyl esters preparation from fresh plant tissues". Analytical Biochemistry, 211:139-143, 1993. This allows for the detection of seeds with modifications in the composition of any fatty acid. Selected seeds showing a desirably high stearic acid content and/or oleic acid and Vmax of thioesterase activity are selected for further breeding purposes. The seeds are cultivated to the fifth generation to form inbreds that have this inheritable and stable genetic trait.

Sunflower oil having a stearic acid content of at least 12% by weight and a oleic acid content of at least 40% by weight, related to the total amount of fatty acids in the oil, is obtainable from sunflower seeds of the invention in any manner known to the person skilled in the art. The sunflower seeds can be converted to crushed seeds or meal before, during or after the oil extraction process. Such extraction methods are well known and for example one method is described in "Bailey's industrial oil and fat products", Vol. 2, Chapter 3; 4th Edition, John Wiley and Sons, New York (1982). The extraction process preferably used does not involve substantial modification of the oil. The extraction process may be selected to avoid substantial chemical, physical or enzymatic rearrangement with no substantial hardening.

In a further embodiment of the method of the invention, the mutation and selection steps may be followed by conventional plant improvement techniques thus leading to seeds having e.g. an even higher stearic acid content or higher oleic acid or reduced linoleic or altered palmitic by weight, or to seeds having a desirable content of one or more other fatty acids. In still another embodiment the seeds of the invention may be subjected to one or more further mutation treatments or additional transformations.

The sunflower plants may be used in breeding programmes for the development of sunflower lines or hybrids, which programmes are aimed at the production of open pollinated or hybrid varieties meeting the requirements of farming practice regarding yield, disease resistance and other agronomically important traits in major sunflower growing areas in the world. Seeds resulting from these programmes may be used in the growing of commercial sunflower crops.

The invention further relates to sunflower plants produced from seeds according to the invention. Thus, the seeds can be used to produce new parent lines that have high stearic acid content and a high oleic acid content in their oil. These lines should also have an increased thioesterase Vmax as indicated in the definition of high levels of thioesterase on the substrate stearyl-ACP at 15 DAF. The seeds may be cultured in the normal way on soil or any other substrate. The production of the modified plants does not require very many additional measures as compared to the growing of ordinary sunflower seeds.

5 The invention also relates to the use in the food industry of sunflower oil extractable from the seeds of the invention. The natural vegetable oil that has been extracted from sunflower seeds of the present invention has a high stearic acid content between 12 and 35% and at least 40 to 80% oleic acid. This oil can be used directly or it can be combined with other oils, in the production of edible fats or fat mixtures, margarine, vegetable-dairy or in the production of confectionery, cosmetic or bakery goods. The present invention is particularly useful in spreads, sauces, ice cream and soups. The oil is also useful for non-edible products such as lipsticks, soap bars, skin and hair oils, lotions and creams.

- 10 In this application the oil of the present invention is interchangeably called oil or fat. The structure of the present invention is in some instances a saturated fatty acid profile that yields a high enough level of saturation to increase the melting point of the oil to make it solid at room temperature. In other saturated fatty acid profiles according to the present invention the material is liquid at room temperature. Both melting points are encompassed by the teachings of the present invention.

- 15 This oil is particularly useful in use as a hardstock material for the formation of spreads. This is a fat that has other oils blended into it. In the preferred use the present invention is used as a hardstock at a level of at least 5% depending on the use of the spread the present invention can be used at levels of at least 20%. Higher levels of the present invention can also be employed depending on the traits desired in the resultant spread. The oil may simply replace oils or fats ordinarily used in this type of products. It is within the reach of the skilled person to determine how to use the oil without performing any inventive labor.

- 20 A number of food products using the oil of the present invention can be used. These food products would include the present oil at levels from 3% to 100% by weight of the other oils (if any) of food product. The advantage of these oils is that they do not have artificial fatty acid isomers, like the 'trans' isomers found in the hydrogenated oils, and, of course, little to no LLD raising cholesterol, like in the animal fats.

- 25 The present invention will be further illustrated by means of the following examples which are given for illustration purposes only and are in no way intended to limit the scope of the invention.

30
35 **EXAMPLES**

The invention also relates to a method for preparing sunflower seeds having an increased stearic acid and oleic acid content as compared to wild type seeds. This method includes the step of treating parent seeds with a mutagenic agent during a period of time and in a

concentration sufficient to induce one or more mutations in the genetic trait involved in stearic acid or oleic acid biosynthesis. This results in an increased production of stearic acid and/ or an increased level of oleic acid. These mutagenic agents include agents such as sodium azide or an alkylating agent, like ethyl methane sulfonate, of course any other

5 mutagenic agent having the same or similar effects may also be used. The treated seeds will contain inheritable genetic changes. These mutated seeds are then germinated and progeny plants are developed therefrom. To increase the traits in the lines the progeny can be crossed or selfed. The progeny seeds are collected and analysed. Seeds having the HOHT trait or the stearic trait can then be crossed to each other to form the HSHO line. Optionally

10 there can be additional cycles of germination, culturing, and selfing to fix the homozygosity of the traits in the lines and crossing and collection of seeds.

Sodium azide and ethyl methane sulfonate were used as mutagenic agents in Example 1 and 2, respectively. Several sunflower lines with a stearic acid content between 12 and 45% have

15 been obtained. In all these cases the original sunflower parent line for the production of the high stearic acid lines used was RDF-1-532 (Sunflower Collection of Instituto de Agricultura Sostenible, CSIC, Cordoba, Spain) that has from 4 to 7% stearic acid content in the seed oil. The high oleic material used herein is derived from the Russian researched oleic lines. It is sufficient to screen oleic lines for a HOHT and use this line for either transformation or for

20 crossing to a high stearic line to develop a HSHO line. A method for the preparations of the stearic lines, and of oleic lines if necessary, have been described in the following examples.

EXAMPLE 1

25 Seeds were mutagenized with a solution of 70 mM of ethyl methane sulfonate (EMS) in water. The treatment was performed at room temperature during 2 hours while shaking (60 rpm). After mutagenesis the EMS solution was discarded and seeds were washed during 16 hours under tap water.

Treated seeds were germinated in the field and plants were self-pollinated. The seeds

30 collected from these plants were used to select new sunflower lines with modifications in the fatty acid composition. By using the method of Garcés, R. and Mancha, M. (supra) the seed fatty acid composition was determined by gas liquid chromatography, after converting the fatty acids into their corresponding methyl esters.

A first plant with 9 to 17% stearic acid content in the oil was selected. The progeny was

35 cultivated for five generations wherein the stearic acid content increased and the new genetic trait became stably fixed in the genetic material of the seed. This line is called CAS-3. The minimum and the maximum stearic acid content of the line were 19 and 35%

respectively. The stearic acid content of oil extracted from seeds from this cell line may thus lie between 19 and 35%.

EXAMPLE 2

5

Sunflower seeds were mutagenized with sodium azide, at a concentration of 2 mM in water. The treatment was performed at room temperature during two hours while shaking (60 rpm). Then the mutagenesis solution was discarded and seeds were washed during 16 hours with tap water.

10

Seeds were planted in the field and plants were self-pollinated. Seeds from these plants were collected, and the fatty acid composition was determined by gas liquid chromatography, after converting the fatty acids into their corresponding methyl esters using the method described in Example 1.

15

Seeds from a plant having around 10% stearic acid in the oil were selected and cultivated for five generations. During this procedure the stearic acid content was increased and the new genetic trait fixed. This line is called CAS-4. A selected sample of this line was analysed resulting in a stearic acid content of 16.1%. The minimum and the maximum values were 12 and 19% respectively.

20

Table 7

Percentage fatty acids				
Line	Palmitic	Stearic	Oleic	Linoleic
25 CAS-3	5.1	26.0	13.8	55.1
CAS-4	5.5	16.1	24.3	54.1

EXAMPLE 3

Sunflower plants were grown from the sunflower seeds of the HOHT line shown in Table 4.

30

Sunflower plants were also grown from the sunflower seeds of CAS-3. The lines were crossed. The plants were assisted by artificially pollination in order to ensure adequate seed production occurred. The F1 seed was produced on the HOHT line, or vice versa, and harvested. The F2 seeds with more than 20% stearate and more than 40% oleate were selected. Although this produces the oil of the present invention the level of production is limited. Therefore fixed inbred lines evidencing seeds with these oil profiles are desirable.

35

These homozygous fixed inbred HSHO lines can then be crossed to form hybrid seed, which will produce F2 seed evidencing the desired oil traits of the present invention. Toward this end the F1 seeds were planted and produced plants were selfed in isolated conditions and

F2 seed was produced. The F2 seed was tested for the three traits, high stearic, high oleic and high levels of thioesterase activity. The remaining portion of the seeds evidencing these traits was employed to grow plants to form F3 seed. The selfing and screening and selection process is repeated to develop the fixed homozygous HSHO line, having the following fatty acid profile, C:16 5.4, C:18.0 24.8, C:18.1 58.5, C:18.2 7.2. Once the trait is fixed similar HSHO lines can cross to form hybrid seed having both traits. According to the invention sunflower plants and seeds from which said oil can be extracted have been obtained by means of a biotechnological process. This high stearic acid content is an inheritable trait and is fairly independent from the growing conditions.

10

EXAMPLE 6

Plants growth conditions.

Sunflower (*Helianthus annuus* L.) seeds from high oleic lines used in example 5 with altered seed fatty acid content was used to test for the thioesterase activities over stearyl-ACP. Plants were cultivated in growth chambers at 25/15°C (day/night) temperature, 16 hours photoperiod and photon flux density of 300 micromol m⁻²s⁻¹. Seeds for analysis were harvested at 15 days after flowering and kept at -20°C.

Radioactive reagents and preparation of acyl-ACPs.

1-14C-Oleic with specific radioactivity of 2.1 GBq/mmol and [9,10(n)-3H] stearic acid with specific radioactivity of 1.9 GBq/mmol were obtained from American Radiolabeled Chemicals Inc. (St.Louis, Mo., USA). To prepare the fatty acid sodium salt, an appropriate volume of fatty acid solution was transferred to a glass tube, the solvent was removed under a stream of nitrogen, and the residue was dissolved in 10% Triton X-100, 0.6 mM NaOH. This solution was heated at 55°C for 1 hour to ensure homogeneity. Acyl-ACPs were prepared using a modification of the enzymatic synthesis procedure of Rock CO, Garwin JL, Cronan, JE Jr. (1981) Preparative enzymatic synthesis of acyl-acyl carrier protein. *Methods Enzymology* 72:397-403. Assays contained 0.1 M Tris-HCl (pH 8.0), 0.4 M LiCl, 5 mM ATP, 10 mM MgCl₂, 2 mM DTT, 130 microM fatty acid sodium salt, 0.27 mM ACP-SH and 1.8 mU of acyl-ACP synthetase (the last two components were purchased from Sigma-Aldrich Quimica S.A. Madrid, Spain) in a final volume of 110 microliter. Reactions were incubated at 37°C for 3 hours. After this time the pH was acidified to 6.0 by adding 1 microliter of 3.6 M HCl and the mixture was cleaned of free fatty acids using a modification of the method described by Mancha M, Stokes GB, Stumpf PK (1975) Fat metabolism in higher plants. The determination of acyl-acyl carrier protein and acyl coenzyme A in a complex lipid mixture 1,2. *Anal. Biochem.* 68:600-608. Consisting in adding an equal volume of isopropanol and washing three times with hexane saturated in water/isopropanol (1: 1; v/v).

Preparation of crude extracts for enzyme assays and protein determination.

Frozen seeds were peeled and grounded in extract buffer containing 20 mM Tris-HCl (pH 8.5), 2 mM DTT and 5% (v/v) glycerol (Dörmann P, Kridl JC, Ohlrogge JB (1994) Cloning and expression in *Escherichia coli* of a cDNA coding for the oleoyl-acyl carrier protein thioesterase from coriander (*Coriandrum sativum* L.). Biochim. Biophys. Acta 1212:134-136) at 1 g of tissues/ 10 ml of buffer. Protein concentrations were measured using a Protein Assay Kit (Bio-Rad) according to the manufacturer's recommendations, with BSA as standard.

Enzyme assays.

Acyl-ACP thioesterase activity was assayed in a final volume of 170 microliter using 130 microliter of crude extract. Control assays had crude extract omitted. Reactions mixtures contained 20 mM Tris-HCl (pH 8.5), 5% glycerol and 2 mM dithiothreitol (DTT) and different concentrations of substrates (stearoyl-ACP and oleoyl-ACP). Incubations were carried out for 20 min at 25°C. Reactions were stopped by the addition of 170 microliter of 1 M acetic acid in isopropanol containing 1 mM of oleic acid. Mixtures were then washed three times with hexane saturated in water/isopropanol (1:1, v/v). Acyl-ACP thioesterase activity was determined by counting the radioactivity of the aqueous phase, which contained the non-hydrolysed substrates. Then, 3 ml of solvent scintillant (purchased from National Diagnostics, Hesse, England) was added and the radioactivity was measured using a scintillation counter (Rackbeta II; LKB, Sweden). Data from acyl-ACP thioesterase assays were fitted to the Michaelis-Menten equation by non-linear least-squares regression analysis using Microcal Origin 4. 1, and correlated to $P < 0.05$, as determined by paired Student's test. V_{max} and K_m were derived from these curves.

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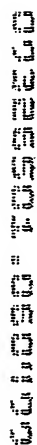
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We claim:

1. Oil from seeds, said oil comprising, an oleic acid content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of said oil.
2. Oil according to claim 1, wherein the oleic acid content is from 55 to 75 wt%.
3. Oil according to claim 1, wherein the stearic acid content is from 15 to 40 wt%.
4. Oil according to claim 1, having a total level of saturated fatty acids of at least 20 wt%.
5. Oil according to claim 1, having a linoleic acid content of less than 20 wt%.
6. Oil according to claim 1 wherein the sunflower oil has at least 90 w/w% of the saturated fatty acid groups in the 1 or 3 position.
7. Oil according to claim 6, wherein from 95 to 100 w/w% of the saturated fatty acid groups are in the 1 or 3 position.
8. Sunflower plant capable of producing seeds according to claim 1.
9. Method of obtaining a sunflower oil having an oleic acid content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of said oil by extracting oil from the seeds according to claim 1.
10. Method according to claim 9, wherein said extraction process does not involve a substantial modification of the sunflower oil.
11. Method according to claim 10, wherein no substantial chemical or enzymatic rearrangement takes place and no substantial hardening.
12. Food product comprising a sunflower oil, said oil having an oleic acid content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of said oil.
13. Food product according to claim 12, wherein the level of sunflower oil is from 3 to 100 wt%.
14. Food product according to claim 13 selected from the group of spreads, sauces, ice-cream, soups, bakery products and confectionery products.
15. Food product according to claim 14, being a spread in which the sunflower oil is used as a hardstock at a level of 5 to 20 wt%.
16. Cosmetic product comprising a sunflower oil, said oil having an oleic acid content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of said oil.
17. Cosmetic product according to claim 16, wherein the level of sunflower oil is from 3 to 100 wt%.
18. Cosmetic product according to claim 17 selected from the group of creams, lotions, lipsticks, soap bars and skin or hair oils.

19. A process for selecting *Helianthus annuus* plants, capable of producing the seeds of claim 1 comprising the steps of:
 - (a) selecting a number of *Helianthus annuus* plants, the mature seeds whereof have a stearic acid content of at least 15 wt% based on the fat present in the seed;
 - (b) selecting a number of *Helianthus annuus* plants, the mature seeds whereof have c;
 - (c) crossing said selected plants of (a) with the selected plants of (b).
20. A process according to claim 19 including the steps of:
 - (d) planting the seeds of the F1 obtained in claim 19;
 - (e) selecting from the F2 progeny those plants which produce seeds having an oleic acid content of more than 40 wt% and a stearic acid content of more than 12 wt% based on the total fatty acid content of said oil.
21. Sunflower meal or crushed seeds obtainable from the seeds according to claim 1.
22. Seeds capable of producing the sunflower oil according to claim 1.
23. An inbred sunflower plant having seeds with oil according to claim 1 including that said seeds have a thioesterase activity over stearyl-ACP of at least 10% of the activity over oleyl-ACP.
24. An inbred sunflower plant having seeds with oil according to claim 23 including a maximum of 10 %, preferably up to 5% w/w of saturated fatty acids in the 2 position of a TAG molecule.



100

Substrate	HOHT (Relative affinity)	HOLT (Relative affinity)
18:0-ACP	~28	~8
18:1-ACP	~98	~98

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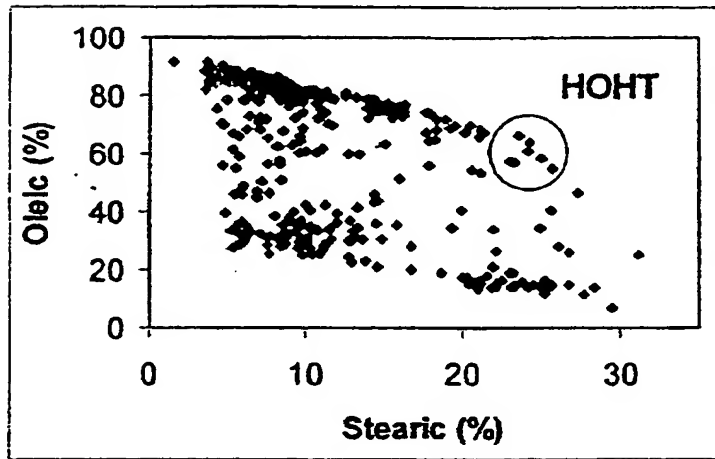
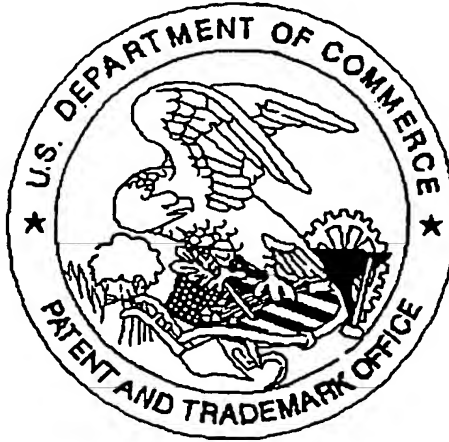


Figure 4.

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